A DISSERTATION ON

Modulatory impact of Millet based bioactive compounds in disrupting the Advance Glycolated End-products (AGE) formation

SUBMITTED TO THE DEPARTMENT OF BIOENGINEERING FACULTY OF ENGINEERING & INFORMATION TECHNOLOGY INTEGRAL UNIVERSITY, LUCKNOW



IN PARTIAL FULFILMENT FOR THE DEGREE OF MASTERS OF TECHNOLOGY IN BIOINFORMATICS

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UNDER THE SUPERVISION OF

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DECLARATION FORM

I, Amna Tamimi, a student of M. Tech Bioinformatics (II Year/ IV semester), Integral University have completed my six months dissertation work entitled "Modulatory impact of millet based bioactive compounds in disrupting the Advanced Glycolated End-products (AGE) formation" successfully from the CSIR- Indian Institute of Toxicology Research, Lucknow under theable guidance of Dr. Parthasarathi Ramakrishnan, Principal Scientist.

I, hereby, affirm that the work has been done by me in all aspects. I have sincerely prepared this project report and the results reported in this study are genuine and authentic.

Amna Tamimi

Course Coordinator



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6th Sept, 2023

To whomsoever it may concern

This is to certify that **Amna Tamimi**, a student of M. Tech. Bioinformatics (II Year/ IV Semester). Integral University, Lucknow has completed a six-month (2nd January 2023 - 1st July 2023) training program under my guidance at the **CSIR-Indian Institute of Toxicology Research.** She has concluded a study on the topic "Modulatory impact of millet based bioactive compounds in disrupting the Advanced Glycolated End-products (AGE) formation". The thesis embodies the results of original work and studies carried out by the student herself and the contents of the thesis do not form the basis for the award of any other degree to the candidate or anybody else from this or any other University/Institution.

During the training, I found her hardworking, enthusiastic and self-motivated. I wish her all the best for future endeavors.

Bert Legand

डी. रॉमोक्स्वान प्रदिसार्त्या /Dr Ramakrishnan Parthasarathi प्रधान वैज्ञानिक / Principal Scientist कम्प्यूटेशनल टॉक्सिकोलॉजी फेरिलिटी Computational Toxicology Facility सेन्टर फोर इनोवेशन एख ट्रान्सलेशनल रिसर्च Cer tre for Innovation and Translational Research तीएसआईआर-भारतीय विषविज्ञान अनुसाधन संस्थान CSIR-Indian Institute of Toxicology Research विषविज्ञान भवन, 31. महात्मा गॉधी मार्ग, लखनऊ-226001, उ.प्र. (भारत) Vishvigyan Bhawan, 31, M.G.Marg, Lucknow-226001, U.P. (INDIA)



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I wish her good luck and bright future.

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TO WHOM IT MAY CONCERN

This is to certify that **Amna Tamimi**, a student of **M. Tech Bioinformatics** (II Year/ IV Semester), Integral University has completed her six months dissertation work entitled **"Modulatory impact of millet based bioactive compounds in disrupting the Advanced Glycolated End-products (AGE) formation**" successfully. She has completed this work from Computational Toxicology Facility, Council of Scientific and Industrial Research – Indian Institute of Toxicology Research (CSIR-IITR), Lucknow under the guidance of **Dr. Parthasarathi Ramakrishnan, Principal Scientist**. The dissertation was a compulsory part of her **M. Tech Bioinformatics**.

I wish her good luck and bright future.

Dr. Alvina Farooqui Professor and Head Department of Bioengineering Faculty of Engineering & Information Technology

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LIST OF ABBREVATIONS

AGE	Advanced Glycation End-products
NF-B	Nuclear transcription factor
ADMET	Absorption, Distribution, Metabolism, Excretion
STAT	Signal Transducer and Transcriptional Activator
TAGE	Toxic Advanced Glycation End-products.
MD	Molecular Dynamics
RAGE	Receptor of Advanced Glycation End-products.
ROS	Reactive Oxygen Species
NADPH	Nicotinamide Adenine Dinucleotide Phosphate.
AD	Alzheimer's Disease
TBI	Traumatic Brain Injury
ALS	Amyotrophic Lateral Sclerosis
GA	Glyceraldehyde
AG	Aminoguanidine
SPC	Simple Point Charge
RMSF	Root Mean Square Fluctuation
RMSD	Root Mean Square Deviation
Rg	Radius of Gyration
BBB	Blood Brain Barrier
HBA	Hydrogen Bond Acceptor
HBD	Hydrogen Bond Donor
B.E.	Binding Energy

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1. Introduction

The advanced glycation end products (AGEs) are organic compounds with a wide range of structural and functional characteristics that are produced in all living organisms [Perrone *et al.*, 2020]. Toxic substances that are formed when protein or fat interacts with sugar in the bloodstream. This process is known as glycation. In other words, AGEs are the irreversible products produced by nonenzymatic glycation, which contain potentially hazardous heterogeneous compounds.

Nonenzymatic glycation occurs when a reducing sugar's reactive carbonyl group comes into contact with proteins, lipids, or nucleic acids [Rungratanawanich W. et al., 2021]. AGE structures come in a wide variety, which points to a wide range of formation methods. In fact, AGEs can be produced from a variety of precursors through a number of different mechanisms, making them both endogenous and exogenous [A.Twarda-Clapa et al., 2022]. The nuclear transcription factor (NF-B), mitogen-activated protein kinase (MAPK), and signal transducer and transcriptional activator (STAT) pathways are only a few of the many signaling pathways that AGEs control. Some metabolic disorders, such as diabetes, atherosclerosis, and Alzheimer's disease, become more severe by the pathogenic function that AGEs play in the development of these conditions [O. Song et al., 2021]. A subclass of AGEs known as toxic AGEs (TAGEs) interacts with the AGE receptors on specific cells and organs to cause harmful consequences [M. Takeuchi et al., 2010]. Toxic AGEs (TAGE) are the name given to these AGEs because they are highly cytotoxic [M. Takeuchi et al., 2020]. When intracellular proteins and glyceraldehyde react, toxic advanced glycation end-products (toxic AGEs, TAGEs) are produced, and their formation is a contributor to a number of cellular diseases.[M. Takeuchi et al., 2022], such as pathogenesis of diabetes mellitus and its related vascular complications, also brain, liver, and heart cells are harmed by TAGE buildup [M. Takeuchi et al., 2020][M. Takeuchi et al., 2016]. Benfotiamine, aminoguanidine, aspirin, tenilsetam, thiazolidinediones, antioxidant agents (carnosine, flavonoids, curcumin), and chelators with AGE inhibition properties (carnosine, pyridoxamine) are just a few of the many substances that have been reported to have a potential role against AGEs. The anti-AGEs efficacy of these substances was shown by preclinical evaluation study results; however, a precise mechanism of action has not yet been identified. Clinical trials are still necessary to confirm many of these drugs' effects [Chen et al., 2018]. Thiazolidinediones have a number of unfavorable side effects, especially when used for an extended period of time. For example, edema, congestive hearts failure, weight gain, fractures, diabetic macular edema, and other conditions [JS Eggleton et al., 202]. Anemia, liver abnormalities, gastrointestinal problems, influenza, lupus, and flu-like symptoms were among the adverse effects of aminoguanidine that were often reported. Considering the toxic and

potential side effects of these synthetic molecules, natural compounds are preferable because of their immense scaffold variety and structural complexity.

Natural remedies have long been available, with many effective medicines coming from plant sources. In the past, natural compounds and their structural analogues have significantly influenced pharmacology, particularly for the treatment of infectious disorders and many chronic diseases like cancer . Currently, several of these natural substances are being researched a spossible medication candidates. Natura lproducts remain to be a possible source of medicine despite all technological improvements made in the creation of synthetic pharmaceuticals because they possess a unique structural diversity that synthetic drugs do not, as compared to ordinary combinatorial chemistry [Dias *et al.*,2012]. They typically have more H-bond acceptors and donors, more sp³ carbon and oxygen atoms than synthetic compounds, higher molecular masses, lower calculated octanol-water partition coefficients, and more rigid molecules which make them distinguishable.

Secondary plant metabolites known as phenolic compounds have been researched for their antioxidant and anti-glycation abilities [MA. Anis *et al.*, 2020]. The natural substance that we work with in the present study as an inhibitor of advanced glycation end products are millets. As millets are rich in nutrients, containing compounds like phenolics that possess anti-inflammatory, antioxidant, and anti-glycation activity, by neutralising carbonyl intermediates during the glycation reaction and chelating transition metals. These millet based bioactive compounds may provide protection by scavenging free radicals, hence preventing the production of AGEs [MA. Anis *et al.*, 2020]. The millet based bioactive compounds were collected from available databases and were subjected to high throughput screening to filter drug-like compounds which structurally mimics the standard drug. Further, the molecular docking was performed for control drugs and derived natural compounds in the binding site of target protein. The validation of the compounds efficacy was done using simultaneous docking with the target in the presence of TAGE. Later, the acquired results were validated by MD simulation study.

Objectives

- 1. The focal point of this study is to use Millet based bioactive compounds to inhibit the formation of Advanced Glycation End (AGE) products. With the help of literature evidence, target protein involved in AGE formation and millet based bioactive compounds were selected.
- 2. Predicting the natural compounds kinetics, drugability, toxicity and pharmacokinetic properties using prediction tools. Pharmacophore analysis, Comparative molecular interaction study and simultaneous docking of controls and natural compounds with the target in presence of TAGE was done.
- 3. Later, MD simulation assesses and analyzes the interaction of hits with the target receptor in comparison to control and TAGE compounds.

2. Review of literature

One can easily distinguish between two types of AGE physiological implications. (1) AGEs encourage the cross-linking of proteins and can serve as molecules that connect various peptide chains in proteins, which can lead to protein aggregation or increased tissue stiffness and a loss of function in both tissues and proteins. This crucial aspect of AGEs' physiological function is frequently explored in relation to ageing, skin conditions, neurological illnesses, diabetes-related cardiovascular disease, and chronic renal disease.(2) Protein-bound AGEs have the ability to directly enhance reactive oxygen species (ROS) formation and have an impact on cell physiology by activating cell membrane receptors such RAGE, OST-48 (OST-48, also known as AGER1), 80 KH protein (AGER2), and galectin-3 (AGER3) [A.J. Lin *et al.*, 2018].

Proteins or lipids that have been nonenzymatically glycated by glucose, or other reducing sugars and their derivatives, such as glyceraldehyde, glycolaldehyde, methylglyoxal, and acetaldehyde, are known as advanced glycation end products (AGEs). A number of diseases, including diabetes and its complications, retinopathy, neuropathy, neurological disorders (such as Parkinson's disease and Alzheimer's disease), atherosclerosis, hypertension, cancers and various other chronic diseases that are influenced by the pathological mechanisms that AGEs perform. Maillard reactions, lipid peroxidation and the polyol pathway are the three main methods of AGE generation [Kuzan *et al.*, 2021].



Figure 1. AGE induced biological effects and co-related diseases. (Lin et al., 2018)

The formation and accumulation of advanced glycation end products (AGEs) in the kidney is worsened by protracted hyperglycemia, dyslipidemia, and oxidative stress. The nonenzymatic Maillard reaction, sometimes known as the "browning reaction," results in AGEs when amino groups from proteins, lipids, or nucleic acids interact with aldehyde groups from sugars and carbonyls. The Amadori product is the first stable byproduct of this reaction. AGEs are created as a result of its subsequent oxidation, fragmentation, rearrangement, and dehydration. The fact that practically all proteins have some AGE changes as a means of indicating molecular age, however, is due to the fact that glucose levels must always be present to support brain function. [MC et al., 2011]. The primary source of vascular oxidative stress in Type 2 diabetes, nicotinamide adenine dinucleotide phosphate (NADPH) oxidase, is expressed and activated at higher levels in human endothelial cells as a result of AGEs.Increased NADPH oxidase activity causes the cellular antioxidants catalase, glutathione, glutathione peroxidase, superoxide dismutase, and glutathione to become depleted, resulting in the production of free radicals.Numerous reactive alpha and beta unsaturated aldehydes are produced by free radicals as a result of the lipid peroxidation of polyunsaturated fatty acids such as glucose are capable of producing advanced lipid peroxidation end products (ALE), which have the potential to serve as glycemic status markers [Dias et al., 2014].

In a hyperglycemic state, the polyol route for glucose metabolism is active, which promotes the generation of AGEs. Aldose reductase, the first and rate-limiting enzyme in this pathway, uses NADPH as the electron donor to catalyze the conversion of glucose into sorbitol. Then sorbitol dehydrogenase converts NAD to NADH and sorbitol into fructose at that point. In the non-enzymatic glycosylation, fructose produced as a byproduct can be phosphorylated into fructose-3-phosphate and subsequently degraded into 3-deoxyglucosone, both of which can be employed as precursors to make AGEs [Kang *et al.*, 2022].

Several diseases are prone to developing or worsening as a result of glycoxidation processes that occur under oxidative stress including diabetes, atherosclerosis, and Alzheimer's disease (AD) as well as in the secondary stages of traumatic brain injury (TBI). The deactivation of numerous enzymes by AGEs, which take the form of intra- and interprotein cross links, accelerates the development of medical condition. Additionally, the downstream inflammatory cascade events are produced by AGEs interactions with their receptors. Particularly in cases of Alzheimer's disease and other neurodegenerative disorders, such as traumatic brain injury (TBI) and amyotrophic lateral sclerosis (ALS), the overexpression of AGE receptors interacts [Reddy *et al.*, 2022].

Further interest in the potential health effects of AGEs was sparked by a number of papers showing an increase in the levels of circulating AGEs in persons with diabetes and chronic renal

disease. [Prasad *et al.*, 2019]. This emerged after a large number of empirical studies showing a connection between AGEs and a number of conditions, including memory loss as we age, the pathophysiology of eye diseases, polycystic ovary syndrome, wound healing, cardiovascular issues, bone health, periodontitis, erectile dysfunction, anaemia in older community-dwelling women, slow walking speed in older adults, peripheral neuropathy, peripheral artery disease, obstructive sleep apnea , islet β -cell dysfunction, cancer, elevated cellular oxidative and inflammatory state, schizophrenia, Alzheimer's disease, and risk for metabolic syndrome in adults and children.

The excessive generation of glyceraldehyde (GA), an intermediary of sugar metabolismin neuronal cells, hepatocytes, and cardiomyocytes is caused by the frequent consumption of high amounts of sugar, which has been linked to the onset/progression of lifestyle-related disease.Toxic advanced glycation end-products (toxic AGEs, TAGE) a subset of AGE, are created when GA reacts with intracellular proteins, accumulating various diseases. Through the receptor for AGEs, the cellular leakage of TAGE impacts the nearby cells, consequently encouraging the development of lifestyle-related diseases.TAGE seeped into the extracellular space, raising the amounts of extracellular TAGE in the circulating fluids, and the intracellular build-up of TAGE caused a variety of cellular diseases [Takeuchiet al., 2021]. Only a few numbers of synthetic AGE inhibitors work by interfering with the first interaction between reducing sugars and amino groups to prevent the development of Schiff bases and AGEs in the glycation process. For instance, acetylsalicylic acid, an ingredient in aspirin, has been shown to block the glycation process by acetylating free amino groups in proteins by preventing the reducing sugars from attaching. Whereas most synthetic inhibitors play a major role in the latter stage of glycation by blocking the formation of Amadori products. A lot of research has been done on aminoguanidine (AG) and pyridoxamine, which are thought to be powerful carbonyl scavengers, as conventional AGE inhibitors. Even though these synthetic compounds show strong and powerful inhibitory effects, they can also cause severe side effects like gastrointestinal disturbance, anaemia, and flu-like symptoms which caused safety concerns that led to the termination of ACTION II clinical trial [Peng et al., 2011].

As natural compounds are comparatively safer than the synthetic compounds for human consumption, it attracted more interests from researchers. These compounds also possess good inhibitory activity against AGE formation and appear to have lesser toxic effects. Recently, there has been an increase in interest in searching natural substances for fresh glycation inhibitors. [*Song Q, Liu J, Dong L, Wang X, Zhang X,* 2021], [*Peng, X., Ma, J., Chen, F., & Wang, M,* 2011]. Polyphenols, polysaccharides, terpenoids, vitamins, alkaloids, and peptides are the six types of

naturally occurring substances that may potentially prevent the production of AGEs based on their structural characteristics. Due to the diversity in the structural and functional unit of natural compounds, they suppress AGE development through a variety of different ways. The seven groups of mechanisms that prevent AGE formation include blocking aldose reductase, protecting protein glycation sites, scavenging oxidative free radicals, controlling AGE receptors, trapping active dicarbonyl compounds, chelating metal ions, and lowering blood sugar levels [*Song Q, Liu J, Dong L, Wang X, Zhang X,* 2021].

Millets are reported to contain phenols with potent anti-crosslinking and anti-glycation effects on collagen along with several biological processes, including those that are anti-oxidative, antihypertensive, anti-tumor, and that prevent the production of AGEs.Additionally, small millets phenolic compounds showed significant suppression of two important postprandial hyperglycaemia-related enzymes (amylase and glucosidase) as well as antioxidant capabilities in several routes [Anis *et al.*, 2020]. Millets may include protein glycation inhibitors that are much better at preventing the production of AGE and protein glycoxidation while also offering advantages over synthetic medicines without having negative side effects.



Figure 2. Health promoting biological activities of bioactive millet . (A.Majid and P. Priyadarshani., 2020)

3. Materials and Methods

3.1 Tools used

RCSB PDB -

RCSB Protein Data Bank (RCSB PDB) (http://www.rcsb.org/) enables advances in science and education by giving access as well as tools for exploration, visualization, and analysis of: experimentally determined 3D structures from the Protein Data Bank (PDB) archive Computed structure models (CSM) from AlphaFold DB and ModelArchive. Examining this data in light of external annotations provides a structural view on biology. The Protein Data Bank (PDB) is a crucial tool for study and instruction in basic biology, health, energy, and biotechnology. The RCSB PDB is the first open access digital data source in biology and medicine and the US data center for the PDB archive. Through an online information portal and a downloadable data archive, PDB provides access to details about the 3D structures of the molecules that make up life. Since the PDB contains a large amount of 3D structure data and has made significant advancements in protein architecture, protein structure prediction is becoming more and more crucial. The procedure has been accelerated using AI and deep learning methods. As a result, it is now able to analyze huge datasets and more quickly find new therapeutic targets. These techniques have been shown to drastically reduce the time and resources needed for protein structure prediction. On top of that, novel protein folds and functions that were previously unknown have been found thanks to the application of AI and deep learning [W. Tian et al., 2018].

PubChem -

A component of the Molecular Libraries Roadmap Initiatives of the US National Institutes of Health (NIH), PubChem (https://pubchem.ncbi.nlm.nih.gov) was established in 2004 as a public archive for data on chemical compounds and their biological activity. The scientific research community can now access a substantial system called PubChem that provides chemical information. Substance, Compound, and BioAssay are three databases that are connected together in PubChem. Both the Compound database and the Substance database store distinct chemical structures that have been taken from the former [S. Kim *et al.*, 2016]. The Substance database houses chemical information that has been deposited by individual data contributors to PubChem. The BioAssay database holds the biological activity information of chemical compounds that have been examined in assay experiments.

SWISS ADME -

Analysing a vast number of molecular structures in order to identify those that have the best chance of treating patients is a step in the process of developing new medications. Pharmacokinetics, which includes the ADME characteristics (Absorption, Distribution, Metabolism, and Excretion) can each be evaluated individually using specialized methodologies, determines how a therapeutic chemical will behave in the body. By estimating early ADME in the discovery phase, the percentage of pharmacokinetics-related failure in the clinical phases is reduced. Computer models have been advocated as an effective substitute for experimental techniques for the prediction of ADME. Chemicals that are probably to be unstable, reactive, poisonous, or interact with biological experiments are removed from chemical libraries using substructure searches and physicochemical parameters. The molecular fingerprint (FP) and the FP2 technique, two molecular descriptors created by cheminformaticians and utilized in classification models for ADME behaviors, are two examples of the molecular descriptors. For the use of machine learning technology, CADD is a pioneering field. An easy way to submit and analyse CADD results is by using the free web tool SwissADME(http://www.swissadme.ch.). It has cutting-edge methods and tools, as well as a number of input options, multi-molecule computing, and the capability to present the findings [H.M. Berman et al., 2000].

Discovery Studio -

The Discovery Studio Visualizer Client is a graphical user interface for accessing Discovery Studio Science. (https://mybiosoftware.com/). This free, user-friendly visualisation tool is the ideal choice for managers and academics who need to engage with modellers but do not need access to the expert-level analytical tools in Discovery Studio [D. Studio., 2008]. It is possible to automate and customise common modelling processes using the Perl-based scripting API, which provides a large selection of common jobs and commands. Various formats that adhere to industry standards are available to view and distribute data on proteins and small compounds. It offers unequalled abilities for collaborating on workflows, sharing data, and utilising computational resources. Additionally, it offers scriptable functions like surface diffraction, chirality and valency checks, secondary structure prediction, access to electrostatics tools, and management of constraints and restrictions. Protein modelling, pharmacophore analysis, and structure-based design may all be easily integrated with one another using the open operating system SciTegic Pipeline PilotScitegic Enterprise Server platformTM, which is the foundation of Discovery Studio.

Autodock Vina -

The inventor and developer of Autodock Vina is Dr. Oleg Trott, of the Molecular Graphics Lab at The Scripps Research Institute (now CCSB). The current version of Autodock Vina is 1.2.0. (https://github.com/ccsb-scripps/AutoDock-Vina). The input and output file formats used by Vina and AutoDock are both PDBOT molecular structure files. PDBOT files can be produced using MGLTools (manually or interactively), and they can also be viewed. Additional files like the AutoDock and AutoGrid parameter files (GPF, DPF), grid map files, and so forth are not required. Vina's design philosophy avoids asking the user to comprehend its implementation specifics, adjust exotic search parameters, cluster results, or have knowledge of complex mathematics (quaternions). All that is required is a description of the search space, including the binding site, and the structures of the molecules being docked. Grid map computation and atom charge assignment are not required. As it can utilise many CPUs or cores to speed up operation, Vina is guicker and more effective than AutoDock 4. Now that numerous ligands may be docked simultaneously in Vina, fragment-based drug design may benefit from this feature. Highthroughput virtual screenings can be accelerated with AutoDock Vina 1.2.0's batch processing and Python bindings. The command line interface or Python can be used to access its simultaneous multiple-ligand docking functionality against a single target structure [J. Eberhardt *et al.*,2021].

With the aid of unbound and MD simulations, homology modelling, and other methods, molecular docking is used to predict the noncovalent binding of macromolecules and ligands. To uncover possible leads for drug development, docking is used to screen digital libraries of compounds that resemble drugs. It is crucial to improve precision while decreasing the amount of time it takes because it uses a lot of computer resources. The need for manually choosing the atom types for grid maps, making grid map files with AutoGrid, choosing the "search parameters," and clustering the results after docking is also eliminated by Vina because it calculates its own grid maps quickly and automatically and does not store them on the disc. It also clusters and ranks the results without providing any intermediate information to the user. [Trott and Olson, 2010].

R Studio –

Visualising and analysing multivariate data are the objectives of principal component analysis. PCA only works with quantitative variables, in contrast to the other approaches. In addition to economics, image processing, healthcare, and security, there are many other applications for principal component analysis in daily life. PCA can help identify important variables that contribute to the variation in data, making it a useful tool for decision-making in a variety of industries. Through R Studio (http://www. rstudio.com), we can create various PCA plots, including Biplot, Screeplot, and Boxplot. Large datasets' dimensionality can also be decreased using PCA, which makes it simpler to analyse and understand complex data [T. RStudio, 2020].

Cygwin –

In order to compile and run Unix or Linux programmes on a Microsoft Windows operating system (OS), Cygwin is a set of free and open-source tools(<u>https://www.cygwin.com/</u>). Users can enjoy a Linux-like experience in a Windows environment thanks to Cygwin [D. Lazenby, 2000]. This functionality makes it simpler for developers to support their applications that operate on the Windows platform and aids in the migration of applications from Unix or Linux-based systems to Windows-based systems.

The dynamic link library (DLL) cygwin1.dll serves as the brains behind the Cygwin suite of tools. The Portable OS Interface, or POSIX, system call capability is provided by the DLL, which acts as an emulation layer. Along with an X server and a comprehensive range of X applications, the Cygwin distribution also includes a sizable number of free utilities, such as the majority of GNU and numerous Berkeley Software Distribution products. Incorporating the words Cygnus and Windows, the term Cygwin was born [C. Xiao *et al., 2016]*.

3.2 METHODOLOGY

3.2.1 Collection of Millet based bioactive compounds

With the help of literature survey structural information about millet based bioactive compounds were obtained. The 92 structure of these compounds were retrieved from PubChem. A dataset of compounds along with their Compound name, PubChem ID, IUPAC name and canonical smiles was prepared. Later these structures were used in the molecular docking studies.

3.2.2 Protein and Ligand preparation

The crystal structure of target protein Aldose reductase (**PDB ID: 4QXA**) was obtained from the RCSB Protein Data Bank (<u>https://www.rcsb.org/structure/4qxa</u>). In order to prepare the target protein and to avoid steric clashes, the structure was pre-processed before docking by removing the heteroatoms, which include water ions, small molecules, and other residues. All the pre-processing is achieved by using Discovery Studio. The chemical structures were obtained from the PubChem (<u>https://pubchem.ncbi.nlm.nih.gov/</u>) database. In addition, 3D conformations of all 94 chemical compounds, including control and phytochemicals were converted from .sdf to .pdb, for further studies.

3.2.3 Virtual Screening

The success of a drug's development process depends on a drug's potency or effectiveness as well as acceptable qualities for absorption, distribution, metabolism, and excretion (ADMET) and toxicity (T). As a result, it aids in the prediction of the compound's drug likeness. In silico ADMET studies were performed on the 92 millet based bioactive compounds with regards to their ADMET, mutagenicity, and carcinogenicity profiles were computed using two tools i.e., Swiss ADME(Daina&A.,2017)and the ADMET module and TOPKAT (TOxicity Prediction by Kobmputer Assisted Technology) module of Biovia Discovery Studio software 4.0.(B. L., &Rohane, 2021)

3.2.4 Pharmacophore Analysis

Pharmacophore have been proven to be successful for the selection of targeted sets of compounds. There are two categories of pharmacophores: structure-based pharmacophores and ligand-based pharmacophore. As we know pharmacophore models are widely used as virtual screening filters, ligand based pharmacophore was done to separate out compounds that have druglike properties. In this study, a pharmacophore model of controls was generated in which only 29 compounds consist of pharmacophoric features such as hydrogen bond acceptors

(HBAs), hydrogen bond donors (HBDs), hydrophobic areas (Hs), aromatic rings (ARs), positively/negatively ionizable groups (PIs/NIs), and exclusion volumes (XVOLs).

3.2.5 Molecular Interaction Analysis

Intermediary procedures were completed using the graphical user interface programme AutoDockTools (**Huey & R., 2012**) including the creation of grid boxes and pdbqt files for protein and ligand preparation (ADT). The prepared file was saved by AutoDock in PDBQT format. Auto Grid was used for the preparation of the grid map using a grid box. The grid size was set to $68 \times 76 \times 70$ xyz points with grid spacing of 0.375 Å and grid center was designated at dimensions (x, y, and z): -12.237, 25.864, -3.284.

The ligand structure is used to generate a scoring grid that will speed up processing. AutoDockVina was used for docking, utilising grid box parameters from the configuration file, protein and ligand information, and so on. The iterated local search global optimizer is used by AutoDockVina. For further examination, the configuration with the lowest binding energy or affinity was extracted and aligned with the receptor structure.

3.2.6 Multiple Ligand Simultaneous Docking

In the present study, an attempt was made on multiple ligand simultaneous docking (MLSD) to mimic the real molecular recognition processes better. Docking multiple substrates simultaneously at the enzymatic sites of PPO would reflect the real biochemical process, gaining mechanistic insights of the concerted action of multiple substrates/cofactors when they approach the binding pockets. Binding dynamics, including possible intermediates and transition states, could even be revealed energetically (Li and Li, 2015).We then validated the binding affinities obtained by docking using, Molecular Dynamics (MD) simulation to cross-check the stability of the complex formed.

3.2.7 Molecular Dynamic Simulations

Molecular dynamic simulation can be performed to check the stability of the complex. It can be done using Gromacs(<u>https://www.gromacs.org/</u>) and will be visualized using UCSF Chimera and Xmgrace to analyze and check the physical movement of the atoms and understand themolecule stable ornotineal system. The classical approach of molecular dynamics (MD) method [Alder and Wainwright, 1957] one simply solves numerically Newton's equations of motion for the interacting many-particle system [Rapaport*et al.*, 2004].

In order to fully comprehend the conformational modifications and dynamic features of the interactions, MD simulations of the strongest protein-ligand complex in comparison to the TAGE

compounds were performed. A total of 4 compounds were selected and simulated using GROMACS 2018 [Loschwitz&Jäckering,2021] (Groningen Machine for Chemical Simulations) with GROMOS96 43a1 force field. The production of ligand parameters related to the selected force field was done using the ATB server. The protein-ligand complexes in the dodecahedron boxes were dissolved using simple point charge (SPC) water molecules, and the system was subsequently neutralised by the addition of counterions. In addition, energy minimization was applied by running the steepest descent minimization integrator at 50000 steps with a force convergence of about 10 kJ/mol. Then, utilising NVT (canonical) and NPT (isothermal-isobaric) ensembles at a temperature of 300 K, all complexes were brought into equilibrium. Thereafter, the systems were simulated for the 10 ns and were analyzed by using different parameters such as Root Mean Square Deviation (RMSD), Root Mean Square Fluctuation (RMSF), Radius Gyration and a number of hydrogen bonds interactions of the protein-ligand complexes etc.

4. Results & Discussions

4.1 Pharmacokinetics and toxicity prediction of ligand molecules

Using the ADMET (Absorption, Distribution, Metabolism, Excretion, and Toxicity) descriptors of Discovery Studio software 4.0 and Swiss ADME, the pharmacokinetic profile of the milletbased bioactive compound was calculated.Mostly in discipline of computer-aided drug design, the discovery of "drug-like" molecules using molecular databases is commonly favoured. The Lipinski's rule of 5 is the most often used drug-likeliness filter. If the violation is 1 or 0, it means that the ligand adheres to the receptor effectively. If the number of violations was greater than two, the compound was eliminated from further consideration. This study revealed that out of 92 compounds, only 48 compounds fulfilled the Lipinski rule of five.



Figure 3. ADMET plot of 2D polar surface area against calculated Alogp98.

The BOILED-Egg method of calculating the penetration through the brain or intestines, as an accurate predictive model, was also used to analyse the link between the lipophilicity and the polarity of the bioactive chemicals. These characteristics are essential when looking for a new

drug, hence almost half of the ligands make strong candidates for being potential medications. The plot is showing green and blue colouredeclipses of 99% confidence limits for intestinal absorption and blood-brain barrier (BBB), as well as red and pink coloured eclipses of 95% confidence limits for intestinal absorption and BBB



Figure 4. Heatmap summary of in-silico ADMET properties of millet based bioactive compounds using R studio.

Figure. 4., shows the summarization of ADMET properties of 92 compounds. The compounds that passed all of the ADMET parameters are shown by the colour red, whereas the compounds that failed all of the parameters are represented by the colour purple.

4.2 Generation and Validation of Ligand-Based Pharmacophore

The controls, ascorbic acid and epigallocatechin gallate were used to construct the pharmacophore. 29 out of 48 millet based bioactive compounds showed reliable pharmacophoric features. These features are identified using colour coded :- Hydrogen bond acceptor (HBA),

green; hydrogen bond donor (HBD), violet; hydrophobic (H), light blue; positive ionizable (pos), red; negative ionizable (neg), blue. The bioactive conformations of the ligands with similar binding modes may be identified using a trustworthy pharmacophore model. The conformation chosen for each compound, which is taken to be the bioactive conformation, corresponds to the conformation that fit the pharmacophore the best. This clearly shows that the filtered 29 compounds have mapped well on the reported pharmacophore model. Thus, verifying its potential for virtual screening.









GLYCITIN



ERIODICTYOL







ARBUTIN

SAPONARIN

NARINGIN







VIOLANTHIN

ONONIN

VITEXIN







RUTIN

GENISTIN





QUERCETIN DIGLUCOSIDE



BETACYANIN

ORIENTIN

Aught.





XYLOTRIOSE

PROCYANIDIN B1

PRODELPHINIDIN



TAXIFOLIN-7-GLUCOSIDE



XYLOTETROSE



LUCENIN -1



TRICIN ETHER

Figure 5. Generated ligand-based pharmacophore model of control Epigallocatechin gallate (65064) with bioactive compounds of millets.



EPICATECHIN GALLATE

LUTEOFEROL

ISOORIENTIN







ISOVITEXIN

ERIODICTYOL

ARBUTIN







SAPONARIN

NARINGIN

VIOLANTHIN



ONONIN



XYLOBIOSE



VITEXIN







SISSOTRIN

RUTIN

GENISTIN







ORIENTIN

QUERCETIN DIGLUCOSIDE

BETACYANIN







PRODELPHINIDIN

TAXIFOLIN-7-GLUCOSIDE

LUCENIN - 1





XYLOTETROSE

TRICIN ETHER

Figure 6. Generated ligand-based pharmacophore model of control Ascorbic acid (54670067) with bioactive compounds of millets.

4.3 Molecular Docking Analysis

A docking study was utilized for better understanding of the structural basis of protein-ligand interactions. The structure of Aldose reductase (PDB ID-4QXA) established by means of X-ray crystallography was used for the docking analysis. The filtered 29 bioactive compounds were docked against the target Aldose reductase (PDB ID-4QXA). The binding energy of these compounds are discussed in the table below.

Table 1.	Binding energy	of docked	ligands and	its active s	ite pocket residues
			-		1

S.No	Compound	CID	Target	Binding	Residue
				Energy	
				(kcal/mol)	
1	Epigallocatechin	65064	4QXA	-8.4 kcal/mol	GLY 19, PHE 32,
	gallate (Control)				GLN 35, LYS 123,
					ASN 124, ALA 133,
					LYS 136
2	Ascorbic acid	54670067	4QXA	-6.3 kcal/mol	VAL 18, GLY 19,
	(Control)				GLY 17,LYS 20, SER
					22, PHE 37,THR 39
3	Taxifolin 7-	14282775	4QXA	-9.7 kcal/mol	SER 21, THR 39, ASP
	glucoside				62,
					THR 63, LYS 123
4	Sissotrin	5280781	4QXA	-9.1 kcal/mol	GLY 17, SER 22,
					ASN 23,
					ASP 33, PHE 37,
					THR 39
5	Rutin	5280805	4QXA	-9.1 kcal/mol	PHE 32, PHE 37,
					LYS 123,ASP 127,
					ALA 133,LYS 136
6	Genistin	5281377	4QXA	-8.9kcal/mol	GLY 17, LYS 31, ASP
					33,PHE 37
7	Ononin	442813	4QXA	-8.8 kcal/mol	LYS20,LYS31,ASP33
8	Luteoforol	114505	4QXA	-8.8 kcal/mol	GLY 19 , PHE 32,
					SER34,GLN35,LYS1
					23,ALA133,LYS136

9	Hesperidin	10621	4QXA	-8.8 kcal/mol	SER 22, ASN 25, LYS
					31,ASP 33, GLN 35,
					PHE 37
10	Prodelphinidin	13831068	4QXA	-8.7 kcal/mol	SER 21, PHE 32, SER
					34,PHE 37, ASP 90,
					LYS 125,ALA 155,
					LYS 156
11	Eriodictyol	440735	4QXA	-8.7 kcal/mol	GLY 19, PHE 32,
					PHE 37,LYS 123,
					ALA 155,LYS 156
12	Glycitin	187808	4QXA	-8.7 kcal/mol	ASP 33, PHE 37,
					THR 39
13	Isoorientin	114776	4QXA	-8.7 kcal/mol	SER21,THR39,LYS12
					3,ALA13
14	Procyanidin B1	11250133	4QXA	-8.6 kcal/mol	LYS 20, SER22,PHE
					37,THR 39
15	Xylotriose	10201852	4QXA	-8.6 kcal/mol	GLY17,PHE32,GLY3
					5, LYS 125, ALA155
16	Hespretin	72281	4QXA	-8.5 kcal/mol	ASP 81, ALA 109,
					VAL 111,LYS 112,
					GLU 113,SER 116,
					THR 173, ASP 177
17	Proanthocyanidins	107876	4QXA	-8.4 kcal/mol	LYS 20, LYS 31, ASP
					33,LYS 123, ALA
					155, LYS 156
18	Tricin ether	101633864	4QXA	-8.3 kcal/mol	GLY17,PHE32,GLN3
					5, LYS 125, ALA 155
19	Betacyanins	6324775	4QXA	-8.3 kcal/mol	VAL 18, GLY 19,GLY
					17,LYS20,SER22,PH
					E37,THR 39
20	Orientin	5281675	4QXA	-8.2 kcal/mol	PHE32,SER34,PHE37
					,GLU113THR 173,
21	Naringin	442428	4QXA	-8.2 kcal/mol	LYS31,ASP33,LYS12
					3,ASN 124, ALA 133,
					LYS 136

22	Isovitexin	162350	4QXA	-8.2 kcal/mol	THR 63, LYS 123
					SER 116, THR 173,
					ASP 177
23	Vitexin	5280441	4QXA	-7.9 kcal/mol	LYS 20, SER 22,
					VAL 18,
					Gly137,Gly138
24	Xylotetrose	101601989	4QXA	-7.7 kcal/mol	LYS 31, ASP 33,
					PHE 37, LYS 125,
					ASP 127, LYS 136
25	Saponarin	441381	4QXA	-7.6 kcal/mol	GLY 19, SER 22, PHE
					37, ASN 124, LYS
					125, ASP 127, LYS
					136
26	Epicatechin gallate	107905	4QXA	-7.5 kcal/mol	GLY 17, GLY 19,ASN
					23, GLN 35 ,PHE 37
27	Violanthin	442665	4QXA	-7.4 kcal/mol	LYS 31, ASP 33
28	Arbutin	440936	4QXA	-7.2 kcal/mol	GLY 17, PHE32,GLN
					35, LYS 125,ALA 155
29	Xylobiose	4439538	4QXA	-6.8 kcal/mol	GLY17,LYS20,SER22
					,ASN 23,PHE37,
					THR 39
30	Lucenin-1	101316994	4QXA	-6.7 kcal/mol	GLU 132, GLN 139,
					ASP 147, PRO 149
31	Quercetin	5282166	4QXA	-6.6 kcal/mol	ASP 81, ALA 109,
	diglucoside				VAL 111, LYS 112,
					GLU 113, SER 116,
					THR 173, ASP 177
		1			1

The compounds Taxifolin-7-Glucoside (14282775) with binding energy (B.E)-9.7 kcal/mol, Sissotrin (5280781) with -9.1 kcal/mol, Rutin (5280805) with -9.1 kcal/mol and Genistin (5281377) with -8.9 kcal/mol showed most promising binding affinity compared to the control (Epigallocatechin gallate (65064) with B.E -8.4 kcal/mol and ascorbic acid (54670067) with B.E -6.3 kcal/mol). The interactions with significant amino acids and the hydrophobic interactions between these molecules and amino acids are potential explanations for the higher binding scores of these compounds. For the purposes of sorting leads, this was taken intoconsideration.



Table 2. Representation of complex formed along with the active site residues.















5.4 Multiple Ligand Simultaneous Docking (MLSD)

The concurrent interactions between the substrate and the macromolecule that are binding together in the presence of an inhibitor are studied using a computer method called multiple ligand simultaneous docking. In this study, MLSD of top 4 potential leads i.e., Taxifolin-7-Glucoside (14282775), Sissotrin (5280781), Rutin (5280805) and Genistin (5281377) against target Aldose reductase (PDB ID-4QXA) along with the TAGEs (Glyceraldehyde (751), Acetaldehyde (177) and Methylglyoxal (880)) was carried out to analyze how these potential leads will perform in the presence of toxic compounds.

Table.3. Binding energy and active site residues of simultaneously docked compounds inpresence of TAGE Glyceraldehyde with Target Aldose reductase (PDB ID-4QXA).

Compound	CID	Complex	Binding	Residue
		Model	Energy	
			(kcal/mol)	
Glyceraldehyde	751	1	-4.65 kcal/mol	GLY19, LYS20, SER
(TAGE 1)				21, THR 39, ASP62,
				THR 63
Taxifolin 7-	14282775	3	-5.21 kcal/mol	GLY 17, GLY19,
glucoside				SER21, ASN25,
				ASP33, LYS 125,
				ASP 127
Sissotrin	5280781	4	-5.65 kcal/mol	GLY16,GLY17,VA18
				,LYS20,SER34,HIS3
				8, LYS 125, LYS 156
Rutin	5280805	8	-1.89 kcal/mol	GLY17, GLN35,
				LEU36, PHE37,
				HIS38, SER88,
				ASP90, ASP91,
				LYS125
Genistin	5281377	6	-5.71kcal/mol	GLY17, LYS20,
				SER22, SER34,
				THR39, ASP62,
				THR63, ALA64,
				GLY65, LYS125
	Compound Glyceraldehyde (TAGE 1) Taxifolin 7- glucoside Sissotrin Rutin Genistin	CompoundCIDGlyceraldehyde751(TAGE 1)14282775glucoside14282775glucoside1Sissotrin5280781Rutin5280805Genistin5281377	CompoundCIDComplex ModelGlyceraldehyde (TAGE 1)7511Taxifolin 7- glucoside142827753Sissotrin52807814Rutin52808058Genistin52813776	CompoundCIDComplexBindingModelEnergy (kcal/mol)Glyceraldehyde (TAGE 1)7511-4.65 kcal/molTaxifolin 7- glucoside142827753-5.21 kcal/molSissotrin52807814-5.65 kcal/molRutin52808058-1.89 kcal/molGenistin52813776-5.71 kcal/mol

Table.4. Binding energy and active site residues of simultaneously docked compounds inpresence of TAGE Acetaldehyde with Target Aldose reductase (PDB ID-4QXA).

S.No	Compound	CID	Complex	Binding	Residue
			Model	Energy	
				(kcal/mol)	
1	Acetaldehyde	177	3	-3.32 kcal/mol	LYS20, GLY19,
	(TAGE 2)				VAL18
2	Taxifolin 7-	14282775	4	-5.21 kcal/mol	GLY17, SER21,
	glucoside				ASN25, ASP33,
					LYS125, ASP127
3	Sissotrin	5280781	2	-5.69kcal/mol	GLY16, GLY17,
					VAL18, LYS20,
					SER34, HIS38,
					LYS125, LYS156
4	Rutin	5280805	6	-1.89 kcal/mol	ASP15, ASP90,
					ASP91, GLN132,
					LYS125
5	Genistin	5281377	3	-5.70kcal/mol	GLY17,LYS20,SER2
					2,SER34,THR39,
					ASP62,THR63,ALA
					64, GLY65, LYS125
1	1	1	1		

Table.5. Binding energy and active site residue of simultaneously docked compounds inpresence of TAGE Methylglyoxal with Target Aldose reductase (PDB ID-4QXA).

S.No	Compound	CID	Complex	Binding	Residue
			Model	Energy	
				(kcal/mol)	
1	Methylglyoxal	880	5	-4.53kcal/mol	GLY16,GLY17,VAL
	(TAGE 3)				18,GLY19,LYS20
2	Taxifolin 7-	14282775	7	-4.95 kcal/mol	LYS20,ASP33,SER3
	glucoside				4,GLN35,PHE37,HI
					S38,ASP90, LYS125
3	Sissotrin	5280781	2	-5.66kcal/mol	GLY16,GLY17,VAL
					18, LYS20, SER 34,
					HIS38,LYS125,LYS1
					56
4	Rutin	5280805	9	-1.87kcal/mol	GLY17,GLN35,LEU
					36,PHE37,SER88,A
					SP90,ASP91,LYS125
5	Genistin	5281377	6	-5.70kcal/mol	GLY17,LYS20,SER2
					2,SER34,THR39,AS
					P62,THR63,ALA64,
					GLY65

Compared to the earlier molecular docking results from this study, it can be deciphered that there have been declining changes in the stability of the molecular interaction and binding energy of the potential leads. The compound Rutin (5280805) was eliminated as it was weakly bound and produced insignificant results. Further validation of the binding affinities obtained by docking will be done by using Molecular Dynamics (MD) simulation to cross-check the stability of the complex formed.





Figure 7. MLSD results of potential leads Taxifolin-7-Glucoside, Sissotrin, Rutin and Genistin with Target Aldose reductase (PDB ID-4QXA) in presence of Glyceraldehyde. (TAGE)





Figure. 8. MLSD results of potential leads Taxifolin-7-Glucoside, Sissotrin, Rutin and Genistin with target Aldose reductase (PDB ID-4QXA) in presence of Acetaldehyde. (TAGE)





Figure. 9. MLSD results of potential leads Taxifolin-7-Glucoside, Sissotrin, Rutin and Genistin with target Aldose reductase (PDB ID-4QXA) in presence of Methylglyoxal. (TAGE)

4.5. Molecular Dynamic Simulation Analysis

The three bound complexes, (Genistin-Target Aldose reductase (PDB ID-4QXA), Sissotrin-Target Aldose reductase (PDB ID-4QXA) and Taxifolin-7-glucoside-Target Aldose reductase (PDB ID-4QXA)) along with two control complexes, (Ascorbic acid-Target Aldose reductase (PDB ID-4QXA) and Epigallocatechin gallate-Target Aldose reductase (PDB ID-4QXA)) and a TAGE complex (Glyceraldehyde-Target Aldose reductase (PDB ID-4QXA)) were subjected to 10 ns all-atom MD simulations. Any compound can cause significant conformational changes when it binds to its Target Aldose reductase (PDB ID-4QXA). The structural dynamics of proteins are estimated using root-mean-square deviation (RMSD).



Figure.10. RMSD plot of complex Glyceraldehyde(TAGE)-Target Aldose reductase (PDB ID-4QXA), Ascorbic acid-Target Aldose reductase (PDB ID-4QXA), Epigallocatechin gallate-Target Aldose reductase(PDB ID-4QXA), Genistin-Target Aldose Reductase (PDB ID-4QXA), Sissotrin-Target Aldose reductase (PDB ID-4QXA) and Taxifolin-7-glucoside-Target Aldose reductase (PDB ID-4QXA).

We computed the RMSD of all the systems to analyze the structural dynamics of target with two control, TAGE and three potential leads. (Figure.10.) the six complex had average RMSD value that lies between 0.25 to 0.255 nm, respectively. Similarly, the average RMSF of two control was 0.133 nm and 0.152 nm and TAGE compound was 0.148, while of genistin, sissotrin and taxifolin-7-glucoside was calculated to be 0.142, 0.154 and 0.137 nm, respectively.



Figure.11. RMSF plot of complex Glyceraldehyde (TAGE)-Aldose reductase, Ascorbic acid-Aldose reductase, Epigallocatechin gallate-Aldose reductase, Genistin-Aldose reductase, Sissotrin-Aldose reductase and Taxifolin-7glucoside-Aldose reductase.



Radius of Gyration

Figure.12. Time evolution of the Radius of gyration.

The radius of gyration (Rg) is a structural variable related to protein's three-dimensional (3D) structure and overall conformation has been used to investigate its compactness and folding tendency (**Figure.12.**). The mean Rg values of the TAGE-Target Aldose reductase (PDB ID-4QXA) were estimated about 1.548 nm, while controls Ascorbic acid-Target Aldose reductase (PDB ID-4QXA) and Epigallocatechin gallate-Target Aldose reductase (PDB ID-4QXA) were estimated about 1.52 nm and 1.492 nm, and potential leads Genistin-Target Aldose reductase (PDB ID-4QXA), Sissotrin-Target Aldose reductase (PDB ID-4QXA) and Taxifolin-7-glucoside-Target Aldose reductase (PDB ID-4QXA) and Taxifolin-7-glucoside-Target Aldose reductase (PDB ID-4QXA) and Taxifolin-7-glucoside radiate (PDB ID-4QXA) packing when binding to potential leads (genistin, sissotrin and taxifolin-7-glucoside) and controls and TAGE represents the loose or tight packaging of protein during the simulation run. run. In this case, no significant structural change was detected in target following binding to compound, where Rg achieves a stable equilibrium state, implying that the complexes are stable along the path.



Figure.13. Intramolecular Hydrogen Bond between Glyceraldehyde (TAGE)- Target Aldose reductase (PDB ID-4QXA) and Ascorbic acid-Target Aldose reductase(PDB ID-4QXA).



Figure.14. Intramolecular Hydrogen Bond between Epigallocatechin gallate-Target Aldose reductase(PDB ID-4QXA) and Genistin-Target Aldose reductase(PDB ID-4QXA).



Figure.15. Intramolecular Hydrogen Bond between Sissotrin-Target Aldose reductase(PDB ID-4QXA) and Taxifolin-7-glucoside-Target Aldose reductase(PDB ID-4QXA).

Within a protein, intramolecular hydrogen bonds (H-bonds) are critical for maintain its 3D structure, overall shape, and to also validate the stability of the complex. Moreover, the number of intermolecular H-bonds of the three potential leads Genistin-Target Aldose reductase (PDB ID-4QXA), Sissotrin-Target Aldose reductase (PDB ID-4QXA) and Taxifolin-7-glucoside-Target Aldose reductase (PDB ID-4QXA) was evaluated to be 2, 4 and 2, respectively (Figure. 14 and Figure. 15). Hence, the higher number of bonds involvement represents the complex stability.

5. Conclusion

Unmasking the previously unidentified bioactive compounds from diverse natural sources has been a focus of research due to the paradigm change toward the development of medications from natural sources. Numerous reports have shown that millet-based bioactive compounds are good natural candidates for blocking various health problems; one of them is Advanced glycation End products (AGE) formation. Advanced glycation End-products (AGEs) are a group of irreversible heterogeneous compounds produced by the breakdown reactions involving reducing sugars and amino acids.At various phases of the glycation process, millet-based bioactive compounds can prevent the production of AGE and protein glycation.

Natural substances have recently gained popularity as superior possible medicines due to their usefulness in boosting health while having fewer negative effects. Given the costly and time-consuming nature of new medication development, pharmaceutical companies face a pressing need to explore new avenues for drug research and development.Pharmaceutical companies will adopt a global trend of using natural medicines as a source of innovative ingredients for pharmaceuticals.

In this study, the focus is on the modulatory impact of those millet-based bioactive compounds that disrupt the AGE formation. The compounds were screened virtually based on ADMET properties and their pharmacophoric models were generated. Further, molecular interaction studies and molecular dynamic simulation have deciphered that genistin, sissotrin, and taxifolin-7-glucoside possess some therapeutic properties that can prevent the formation of AGE products. This has been established depending upon the binding of the bioactive compounds with the protein target as well as the stability of formed complexes. These compounds inhibit thiol group oxidation, conformational changes, and structural modification in target aldose reductase (PDB ID-4QXA) by scavenging free radical, metal chelation activities, capturing active carbonyl compounds, covering the glycation sites of proteins, and lowering blood glucose levels.

In conclusion, research indicates that millet-derived bioactive compounds offer an appropriate framework and a broader field of investigation for the discovery of potential AGE inhibitors.

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